77. Chiral [²H]-Labelled Methylene Groups in Trienoic- and Dienoic Fatty Acids: a Facile Approach *via* Asymmetric Epoxidation of [²H]Allyl Alcohols

by Christoph Neumann and Wilhelm Boland*

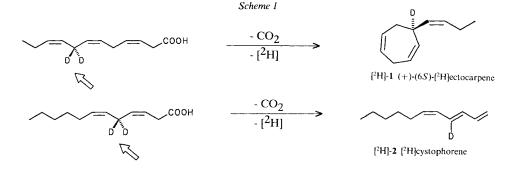
Institut für Organische Chemie der Universität, Richard-Willstätter-Allee 2, D-7500 Karlsruhe

(28.II.90)

Chiral [²H]-labelled methylene groups flanked by two double bonds within (poly)unsaturated fatty acids are readily available from *trans*-2,3-epoxy[2,3-²H₂]alk-4-yn-1-ols, obtained in their turn by asymmetric epoxidation of the corresponding (*E*)-[2,3-²H₂]alk-2-en-4-yn-1-ols (see *Scheme 3*). The procedure is exemplified for (8S,3Z,6Z,9Z)-[7,8-²H₂]trideca-3,6,9-trienoic acid ((8S)-11) and (8*R*)-11 (*Scheme 4*) as well as for (5S,3Z,6Z)-[4,5-²H₂]deca-3,6-dienoic acid ((5S)-13) and (5R)-13 (*Scheme 5*).

1. Introduction. – Unsaturated fatty acids possessing a chiral methylene group (by isotopic substitution with ²H or ³H) have been successfully employed for many times to evaluate the stereochemical course of enzymatic reactions of the lipid metabolism. In particular, the unique physiological activities of the products of lipoxygenases and cyclooxygenases have given an enormous impact to follow their transformations with chirally labelled C_{18} - or C_{20} -fatty acids. Besides a deeper insight into the mechanistic precision work of enzymes, the stereochemical approach also provides a reliable basis for the classification of reaction types and the conceptual description of less well understood biosynthetic pathways.

In continuation of our studies on the biosynthesis of algal pheromones like, *e.g.*, ectocarpene 1 or cystophorene 2 from dodecatrienoic or dodecadienoic acid, such a stereochemical approach seemed to us particularly promising. In a previous model study with the flowering plant *Senecio isatideus*, we have already shown that the biosynthesis of 1 starts from (3Z,6Z,9Z)-dodeca-3,6,9-trienoic acid and proceeds *via* oxidative decarboxylation and loss of a single H-atom from C(8) of the precursor acid [1] (*Scheme 1*). The linear hydrocarbon 2 is a metabolite of (3Z,6Z)-dodeca-3,6-dienoic acid, but besides



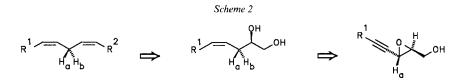
decarboxylation, in this case, a single H-atom from the methlyene group at C(5) is involved [1]. To facilitate the mass-spectroscopic detection and analysis of the labelled metabolites, these studies have been carried out with [²H]-labelled (3Z,6Z,(9Z))-undeca-3,6,(9)-di(tri)enoic acids or (3Z,6Z,(9Z))-trideca-3,6,(9)-di(tri)enoic acids. The unnatural acids are metabolized in the same way as the genuine precursors, but their metabolites can be easily analyzed by MS without being superimposed by the plant's own ¹H-compounds.

Using now chiral [²H]-labelled (3Z,6Z,9Z)-trideca-3,6,9-trienoic acids (²H at C(8)) or (3Z,6Z)-deca-3,6-dienoic acids (²H at C(5)) of known absolute configuration and enantiomeric excess, the site specificity of the enzyme(s) (*viz* loss of the (*pro-S*)- or (*pro-R*) H-atom from the methylene groups at C(8) or C(5)) might be simply assessed by looking onto the metabolic fate of the individual hydrogen isotopes *via* MS analysis of the products. In conjunction with the already known absolute configuration of the major product, namely (6S)-ectocarpene (1) [1] [2], further information on the folding and cyclization of the fatty acid at the active center of the enzyme(s) may be also gained by this approach.

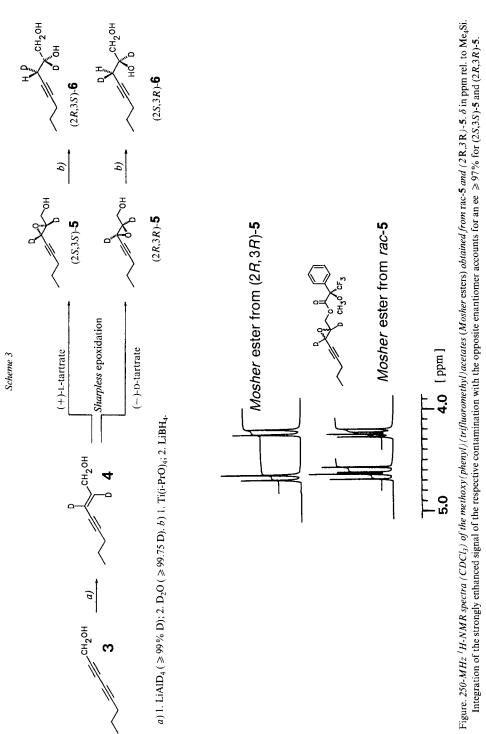
Various syntheses of chiral [²H]- or [³H]-labelled polyenoic fatty acids have appeared in the literature over the past ten years. Many of them make use of a final microbial desaturation of chiral [²H]- or [³H]-labelled stearic or icosanoic acids to α -linolenic or arachidonic acids, respectively [3]. Although this method is straightforward and effective, it is, of course, not applicable to the synthesis of unnatural and short-chain fatty acids. Thus, for the preparation of chiral (3Z,6Z,9Z)-[²H]trideca-3,6,9-trienoic acids, only a highly flexible total synthesis, like *e.g. Corey*'s route towards (7*R*)-[7-²H]arachidonic acid (*ex* tartaric acids [4]) can be successfully employed. However, the multitudiny of laborious steps leading to simple chiral intermediates has to be considered as a major drawback of this approach.

We, therefore, designed and elaborated a new route towards chiral $[^{2}H]$ -labelled methylene groups of unsaturated fatty acids which combines the advantages and simplicity of the acetylenic chemistry and the procedural ease of the *Sharpless* asymmetric epoxidation with the general strategy of the *Corey* approach.

2. Highly Enantiomerically Pure (8R,3Z,6Z,9Z)- and (8S,3Z,6Z,9Z)-[7,8-²H₂]-Trideca-3,6,9-trienoic Acids via Asymmetric Epoxidation. – According to the general concept outlined in *Scheme 2*, 2,3-epoxyalk-4-yn-1-ols are ideally suited for the construction of chiral methylene groups which are flanked by two double bonds. The 2,3-epoxy

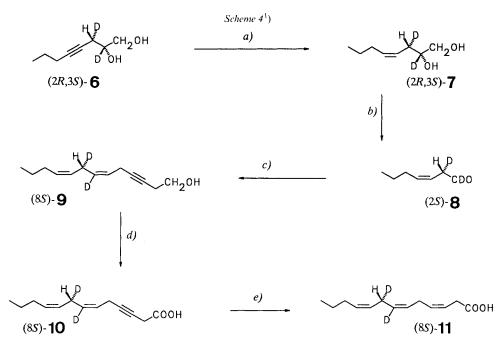


alcohols can be regioselectively reduced at C(3) at which a H-atom or one of its isotopes is introduced with inversion of the configuration at this chiral center. Subsequent cleavage of the resulting diol provides an aldehyde which in turn may be olefinated to give the C-backbone of the desired fatty acid. Since the starting alkynols as well as the *Wittig* reagents may contain various alkyl moieties, additional double or triple bonds, as well as



certain functional groups, the current concept can be easily extended to the synthesis of a wide range of chiral (²H)polyenoic fatty acids.

For the synthesis of (3Z, 6Z, 9Z)-trideca-3,6,9-trienoic acid, labelled with ²H at C(8), octa-2,4-diyn-1-ol (3) [5] is the most convenient starting material (Scheme 3). Using the commercially available highly enriched LiAl²H₄ ($\ge 99\%$ ²H), the required ²H-atom can be introduced with stereospecific reduction of the first triple bond to give the (E)-alcohol 4. The introduction of a second ²H-atom by hydrolysis of the organoaluminum intermediate with ${}^{2}H_{2}O$ ($\ge 99\% {}^{2}H$) is convenient, since it places a permanent ${}^{2}H$ -atom onto the C-backbone of the acid (8S)-11 or (8R)-11 (see below) which is not tackled by the enzymatic conversion of the precursor into ectocarpene 1 [1] (cf. Scheme 1). According to Vasella and coworkers, the asymmetric epoxidation of 4 is readily achieved with the system Ti(t-BuO), and (-)-D- or (+)-L-diethyl tartrate in CH₂Cl₂ at -30° [6]. The use of $Ti(t-BuO)_4$ is essential, since the more commonly employed $Ti(i-PrO)_4$ effects nucleophilic ring opening of the very reactive acetylenic epoxides (2S,3S)- and (2R,3R)-5. As can be shown by 'H-NMR analysis of their Mosher esters [7], both epoxides are of high enantiomeric purity ($\geq 97\%$ ee; Fig.). Application of the more recently developed catalytical variant of the asymmetric epoxidation [8] is possible, but the ee of the product does not exceed 90% [9]. Following complexation of (2S,3S)- or (2R,3R)-5 with $Ti(i-PrO)_4$, the epoxides are regioselectively reduced at C(3) using LiBH₄ [10] to give the two diols (2R,3S)- and (2S,3R)-6, respectively.

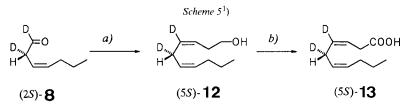


a) Nickel boride/H₂. b) NaIO₄. c) 1. ThpOCH₂CH₂C \equiv CCH₂CH=PPh₃ [4]; 2. pyridinium *p*-toluenesulfonate. d) CrO₃/H⁺. e) Lindlar's catalyst/H₂.

¹) The same reactions were performed with the corresponding enantiomers (see *Exper. Part*).

Since β , γ -olefinic aldehydes are less prone to enolization (*viz* loss of ²H and chirality) than their acetylenic analogoues, the triple bond of (2*R*,3*S*)-**6** is next reduced with nickel boride/H₂ [11] to yield the olefinic diol (2*R*,3*S*)-**7** (Scheme 4)¹). Subsequent cleavage of the diol moiety with NaIO₄ proceeds under neutral conditions and affords the aldehyde (2*S*)-**8** as key intermediate for the synthesis of chiral (²H)trienoic and (²H)dienoic acids. To avoid racemization and isomerization of the sensitive β , γ -unsaturated (²H)aldehyde (2*S*)-**8**, the crude product is immediately converted to (8*S*)-**9** by *Wittig* reaction and deblocking (Py/TsOH in MeOH), following the protocol of *Corey* and *Lansbury* [4]. Oxidation with CrO₃/H⁺ furnishes the acetylenic acid (8*S*)-**10**, and a final hydrogenation with *Lindlar*'s catalyst/H₂ yields the chiral [²H]acid (8*S*)-**11** without touching the chiral center C(8) (\geq 97% ee; secured by exclusive loss of either ¹H or ²H on administration of (8*S*)-**11** and (8*R*)-**11** to the flowering plant *Senecio isatideus* [12], *vide infra*).

3. (5R,3Z,6Z)- and (5S,3Z,6Z)-[4,5- $^{2}H_{2}]$ Deca-3,6-dienoic Acids. – The (3Z,6Z)-[4,5- $^{2}H_{2}]$ deca-3,6-dienoic acids, chirally labelled at C(5), are readily accessible from the aldehydes (2S)-8 and (2R)-8 by an analogous sequence. Treatment of (2S)-8 with $\{3$ -[(tetrahydropyran-2-yl)oxy]propylidene $\}$ triphenylphosphorane affords the tetrahydropyranyl ether which is converted into the free alcohol (5S)-12 on treatment with Py/TsOH in MeOH (*Scheme 5*)¹). Final oxidation with CrO₃/H⁺ gives the decadienoic acid (5S)-13 in 37% overall yield from (2S)-8.



a) 1. ThpOCH₂CH₂CH=PPh₃; 2. pyridinium p-toluenesulfonate. b) CrO₃/H⁺.

Administration experiments with (8S)-11 or (8R)-11 and freshly disconnected sprouts of *Senecio isatideus* were already successful. Thus, (8S)-11 is converted to ecto-carpene 1 with retention of the ²H-label, while (8R)-11 is transformed into 1 with complete loss of the ²H-atom from C(8) of the precursor acid. The detailed study is published elsewhere [12].

Financial support by the *Deutsche Forschungsgesellschaft* and the *Fonds der Chemischen Industrie*, Frankfurt am Main, is gratefully acknowledged. We also thank *Bayer AG*, Leverkusen, and *BASF*, Ludwigshafen, for generous supply with solvents and chemicals.

Experimental Part

General. Highly enriched LiAl²H₄ ($\ge 99\%$ ²H) was from *Fluka*, Buchs, Switzerland, and ²H₂O (99.75% ²H) from *E. Merck*, Darmstadt, FRG. Reactions were performed under Ar. Solvents and reagents were purified and dried prior to use. Anh. MgSO₄ was used for drying operations. Solns. were usually concentrated by flash evaporation under reduced pressure. Anal. GLC: *Carlo Erba* gas chromatograph, *HRGC 5300*, *Mega* series, equipped with fused-silica capillaries, *SE 30* (10 m × 0.31 mm); H₂ at 30 ml/s served as carrier gas. Polarimetry: *Perkin-Elmer 141*, optical rotations at 21°. IR (cm⁻¹): *Perkin-Elmer-882* IR spectrophotometer. ¹H-NMR (250 or 400 MHz; CDCl₃, TMS as internal standard): *Bruker Cryospec WM 250* and *Bruker WM 400*. MS (*m/z*): *Finnigan MAT 90* GLC/MS system and *Finnigan ITD 800* combined with a *Carlo Erba* gas chromatograph, model *Vega*, equipped with a fused-silica capillary, *OV 101* (10 m × 0.32 mm); He at 30 ml/s as carrier gas.

(E)-[2,3- ${}^{2}H_{2}]Oct$ -2-en-4-yn-1-ol (4). A soln. of 3 (1.23 g, 10.0 mmol) in dry THF (20 ml) is slowly added with cooling (0°) to a well-stirred suspension of LiAl²H₄ ($\ge 99\%$ ²H; 0.355 g, 8 mmol) in THF (50 ml). After 4 h, the cold mixture is hydrolyzed by addition of ${}^{2}H_{2}O$ ($\ge 99\%$ ²H; 10 ml), and the product is extracted with Et₂O (5 × 50 ml). Pure 4 is obtained after drying, evaporation, and chromatography on silica gel (pentane, then pentane/Et₂O 4:1): 1.14 g (90%). Colorless oil. IR (film): 3337s (br.), 2967s, 2936s, 2876s, 2237w, 1604w, 1462m, 1431m, 1379m, 1338m, 1327m, 1227w, 1173w, 1095s, 1045m, 999s, 878w, 793w, 708s, 679m. ¹H-NMR (CDCl₃): 4.21 (s, 2 H–C(1)); 2.32 (t, 2 H–C(6)); 2.25 (br. s, OH); 1.49 (sext., 2 H–C(7)); 0.92 (t, 3 H–C(8)). MS (70 eV): 109 (7, M^{+-} OH), 97 (100), 83 (12), 80 (7), 69 (45), 55 (10), 40 (18). HR-MS: 126.1009 (C₈H₁₀²H₂O, M^{++} , calc. 126.1013).

 $(2S,3S)-2,3-Epoxy[2,3-^2H_2]oct-4-yn-1-ol ((2S,3S)-5)$. A soln. of (+)-L-diethyl tartrate (6.4 g, 31 mmol) in CH₂Cl₂ (10 ml) is slowly added with stirring to a cold (-25°) soln. of Ti(*t*-BuO)₄ (10.3 g, 30.4 mmol) in CH₂Cl₂ (30 ml). The complex is aged for 30 min at -25°, before **4** (2.0 g, 16 mmol) in CH₂Cl₂ (60 ml) and *t*-BuOOH (12.2 ml, 26.4 mmol; 3M in 2,2,4-trimethylpentane) are sequentially added. The temp. is lowered to -30°, and after 5 h, the mixture is hydrolyzed by addition of aq. tartaric acid (150 ml of a 10% soln.). The product is extracted with Et₂O (5 × 100 ml), washed with aq. tartaric acid (10%) and brine, dried, and evaporated. Chromatography on silica gel (pentane/Et₂O 7:3) affords 1.1 g (55%) of a colorless oil. IR (film): 3429s (br.), 2967s, 2937s, 2876s, 2250m, 1461m, 1429m, 1381m, 1339m, 1276w, 1228w, 1164w, 1084s, 1030s, 964m, 940m, 895m, 871m, 800m, 769m, 759m, 745m. ¹H-NMR (CDCl₃/²H₂O, 250 MHz): 3.95 (d, J = 12.5, 1 H-C(1)); 3.69 (d, J = 12.5, 1 H-C(1)); 2.19 (t, 2 H-C(6)); 1.55 (sext., 2 H-C(7)); 0.99 (t, 3 H-C(8)). MS (70 eV): 112 (9 M⁺ - CH₂O), 109 (10), 98 (50), 97 (70), 80 (100), 78 (42), 69 (71), 66 (50), 63 (18), 55 (72), 52 (66), 44 (66). HR-MS: 142.0962 (C₈H₁₀²H₂O₂, M⁺, calc. 142.0960).

(2R,3R)-2,3- $Epoxy[2,3-^2H_2]oct$ -4-yn-1-ol ((2R,3R)-5). Prepared from 4 (2.0 g, 16 mmol) and (-)-D-diethyl tartrate as described for (2S,3S)-5. Yield: 1.1 g (55%). Spectroscopic data: identical with those of (2S,3S)-5.

Mosher Esters of the 2,3-Epoxy Alcohols: General Procedure [7]. A soln. of (2S,3S)-, (2R,3R)-, or (2RS,3RS)-5 (20 µl, 0.20 mmol), (+)-(S)-methoxy(phenyl)(trifluoromethyl)acetyl chloride (60 µl, 0.24 mmol), and 4-(dimethylamino)pyridine (35 mg, 0.28 mmol) in CH₂Cl₂ (600 µl) is stirred for 30 min. The solvent is evaporated, the amine precipitated by pentane, and the pure *Mosher* ester (45 mg, 95%) obtained after chromatography on silica gel (pentane/Et₂O 9:1). IR (film; identical for both *Mosher* esters): 3072w, 2967m, 2853w, 2250w, 1757s, 1495w, 1451m, 1401w, 1381w, 1353w, 1324w, 1271s, 1239s, 1170s, 1123s, 1082m, 1017s, 973m, 914w, 886w, 797w, 764m, 718s, 697m, 641m. ¹H-NMR (CDCl₃, 400 MHz; *Mosher* ester of (2R,3R)-5): 7.46 (m, 5 arom. H); 4.68 (d, J = 12.2, 1 H–C(1)); 4.23 (d, J = 12.2, 1 H–C(1)); 3.56 (d, J = 1.1, MeO); 2.17 (t, 2 H–C(6)); 1.52 (sext., 2 H–C(7)); 0.97 (t, 3 H–C(8)). ¹H-NMR (*Mosher* ester of (2S,3S)-5): identical with that of (2R,3R)-5, except for 4.61 (d, J = 12.2, 1 H–C(1)); 4.27 (d, J = 12.2, 1 H–C(1)).

(-)-(2R,3S)- $[2,3-^{2}H_{2}]Oct-4$ -yne-1,2-diol ((2R,3S)-6). To a soln. of (2S,3S)-5 (2.8 g, 19.7 mmol) in dry THF (200 ml) is slowly added with stirring at r.t. Ti(i-PrO)₄ (7.32 g, 29.5 mmol). After 5 min, LiBH₄ (1.64 g, 75.4 mmol) is added in three portions, and stirring is continued for 5 h at 30°. The mixture is hydrolyzed at 0° with H₂SO₄ (250 ml, 5% soln.), the product extracted with Et₂O (5 × 100 ml), and the combined org. layer washed with H₂O. After drying and evaporation, chromatography on silica gel (pentane/Et₂O 6:4) yields 1.5 g (53%) of (2R,3S)-6. Colorless viscous oil. [α]_D = -10.14 (CH₂Cl₂, c = 7.49). IR (film): 3371s (br.), 2964s, 2935s, 2875m, 2159w, 1632w, 1490m, 1438s, 1380s, 1338s, 1281m, 1167w, 1134m, 1109m, 1037s, 950w, 912w, 884w, 849w, 796w. ¹H-NMR (CDCl₃, 400 MHz): 3.73 (d, J = 11.2, 1 H-C(1)); 3.58 (d, J = 11.2, 1 H-C(1)); 2.46 (s, OH); 2.37 (t, J = 2.2, 1 H-C(3)); 2.2 (s, OH); 2.13 (dt, J = 7.3, 2.2, H-C(6)); 1.50 (sext., J = 7.3, 2 H-C(7)); 0.96 (t, 3 H-C(8)). MS (70 eV): 126 (16, M⁺ - H₂O), 115 (29, 113 (23), 101 (17), 97 (25), 83 (41), 82 (34), 68 (100), 67 (84), 62 (63), 55 (53). HR-MS: 126.1008 (C8H₁₂²H₂O₂, M⁺ - H₂O, calc. 126.1013).

(+)-(2S,3R)-[2,3- $^{2}H_{2}]Oct$ -4-yne-1,2-diol ((2S,3R)-6). Prepared from (2R,3R)-5 (2.8 g, 19.7 mmol) as described for (2R,3S)-6. Yield: 1.5 g (53%). [α]_D = +9.5 (CH₂Cl₂, c = 5.9). Spectroscopic data: identical with those of (2R,3S)-5. HR-MS: 126.1000 (C₈H₁₂²H₂O₂, M⁺⁺ - H₂O, calc. 126.1013).

(-)-(2R,3S,4Z)-[2,3- $^{2}H_{2}]Oct$ -4-ene-1,2-diol ((2R,3S)-7). A well stirred suspension of Ni(AcO)₂ (80.0 mg, 0.32 mmol) in EtOH (20.0 ml) is reduced in a closed system, connected to a gas burette and flushed with H₂, by slow injection of NaBH₄ (0.31 ml, 1M in EtOH). The active catalyst is poisoned by ethane-1,2-diamine (40 µl), and (2R,3S)-6 (3.6 g, 25 mmol) is injected. When the H₂ uptake ceases, the catalyst is filtered off, and Et₂O (100 ml) is added. The org. layer is washed with 5% HCl soln. (2 × 10 ml) and H₂O (10 ml). After drying and evaporation, chromatography on silica gel (pentane/Et₂O 6:4) affords 3.5 g (95%) of (2R,3S)-7. Colorless viscous oil. [α]₀ = -6.93 (c = 4.56). IR (film): 3394s (br.), 3012m, 2961s, 2934s, 2874s, 2152w, 1657w, 1456m, 1402m, 1378m, 1337w, 1275m, 1153m, 1109m, 1041s, 982w, 950m, 880w, 849m, 796w. ¹H-NMR (CDCl₃, 400 MHz): 5.36-5.59 (m, 2 olef. H); 3.65 (d, J = 11.2, 1 H–C(1)); 3.46 (d, J = 11.2, 1 H–C(1)); 2.42 (s, 1 OH); 2.17 (d, J = 6.6, 1 H–C(3)); 2.02 (q, 2 H–C(6)); 1.67 (s, 1 OH); 1.37 (sext., 2 H–C(7)); 0.89 (t, 3 H–C(8)). MS (70 eV): 146 (2, M^+), 128 (36), 115 (16), 98 (12), 97 (78), 85 (79), 69 (43), 62 (100), 56 (60). HR-MS: 146.1255 (C₈H₁₄²H₂O₂, M^+ , calc. 1146.1275).

(+)-(2S,3R,4Z)- $[2,3^{-2}H_2]$ Oct-4-ene-1,2-diol ((2S,3R)-7). Prepared from (2S,3R)-6. (2.8 g, 19.7 mmol) as described for (2R,3S)-7. Yield: 1.5 g (53%). [α]_D = + 7.18 (CH₂Cl₂, c = 4.65). Spectroscopic data: identical with those of (2R,3S)-7. HR-MS: 146.1239 (C₈H₁₄²H₂O₂, M⁺, calc. 1146.1275).

(8S,6Z,9Z)-[7,8-²H₂]Trideca-6,9-dien-3-yn-1-ol ((8S)-9). At 0°, (2R,3S)-7 (0.6 g, 4.0 mmol) is cleaved by stirring with NaIO₄ (0.97 g, 4.5 mmol) in H₂O (20 ml). After 1.5 h, the resulting (2S)-8 is extracted with Et₂O $(5 \times 20 \text{ ml})$, and the combined org. layers are dried (Na₂SO₄). To avoid losses of the volatile (2S)-8, the bulk of solvent is removed at 200 Torr, and the crude aldehyde is then immediately transferred into a cold (-78°) soln. of {6-[(tetrahydro-2*H*-pyran-2-yl)oxy]hex-3-ynylidene}triphenylphosphorane (8.2 mmol; prepared from the corresponding phosphonium iodide by addition of 1.0 equiv. of BuLi) in dry THF (100 ml) [4]. The mixture is allowed to come to r.t. and hydrolyzed with sat. aq. NH₄Cl soln. (100 ml), and the product is extracted with pentane. The combined org. layers are washed with H₂O, dried, and evaporated, and the Thp ether is prepurified by chromatography on silica gel (pentane/Et₂O 9:1). The compound is deblocked in refluxing MeOH (150 ml) in the presence of pyridinium p-toluenesulfonate (100 mg). After 3 h, the alcohol is isolated as usual, and chromatography on silica gel (pentane/Et₂O 4:1) yields 0.5 g (63% overall) of (8S)-9 as a colorless oil. IR (film): 3346s (br.), 3013m, 2962s, 2933s, 2874s, 2237w, 2156w, 1636w, 1607w, 1595w, 1465m, 1399m, 1377m, 1334m, 1276m, 1103m, 1046s, 928w, 880w, 848w. ⁱH-NMR (CDCl₃, 400 MHz): 5.45–5.31 (m, 3 olef. H); 3.64 (t, 2 H–C(1)); 2.92 (td, J = 6.5, 2.4, 2 H-C(5); 2.73 (br. s, 1 H–C(8)); 2.42 (tt, J = 5, 2.4, 2 H–C(2)); 2.03 (q, 2 H–C(11)); 1.58 (s, OH); 1.39 (sext., 2 H) 2 H-C(12)); 0.88 (t, 3 H-C(13)). MS (70 eV): 194 (0.5, M⁺), 63 (5), 149 (9), 147 (11), 133 (15), 119 (53), 107 (52), 93 (100), 92 (84), 81 (78), 80 (65), 69 (61), 55 (45), 53 (28), 51 (32), 41 (92), 39 (82). HR-MS: 194.1591 (C₁₃H₁₈²H₂O, M^{+} , calc. 194.1639).

 $(8R,6Z,9Z)-[7,8-^2H_2]$ Trideca-6,9-dien-3-yn-1-ol ((8R)-9). Prepared from (2S,3R)-7 (0.6 g, 4.0 mmol) as described for (8S)-9. Yield: 0.5 g (63%). Spectroscopic data: identical with (8S)-9.

(8S,6Z,9Z)- $[7,8-^{2}H_{2}]$ Trideca-6,9-dien-3-ynoic Acid ((8S)-10). A soln. of CrO₃ (0.74 g, 7.4 mmol) in H₂O (2.1 ml) and conc. H₂SO₄ soln. (0.675 ml) is added slowly with stirring at 0° to a soln. of (8S)-9 (0.48 g, 2.5 mmol) in acetone (35 ml). When the red color of the oxidant persists, the soln. is stirred for 1.5 g at 0° without further addition of Cr(VI). H₂O (60 ml) is added, and the green soln. is extracted with Et₂O (5 × 50 ml). The combined org. layers are washed with brine, until the aq. phase remains colorless. After drying, evaporation and chromatography on silica gel (pentane/Et₂O 7:3), 0.322 g (64%) of (8S)-10 are obtained. IR (film): 3600–3200s (br.), 3014s, 2963s, 2935s, 2876s, 2660w, 2558w, 2243w, 1720s, 1455w, 1421m, 1399m, 1284m, 1228s, 1178m, 1103w, 928w. ¹H-NMR (CDCl₃)²H₂O, 400 MHz): 5.44–5.39 (m, 3 olef. H); 3.33 (t, *J* = 2.4, 2 H–C(2)); 2.99 (td, *J* = 6.9, 2.4, 2 H–C(5)); 2.78 (br. s, 1 H–C(8)); 2.03 (q, 2 H–C(11)); 1.37 (sext., 2 H–C(12)); 0.91 (t, 3 H–C(13)). MS (methyl ester; 70 eV): 223 (5, M^{+} + 1), 195 (6), 181 (6), 165 (12), 163 (13), 149 (31), 148 (26), 147 (23), 133 (57), 123 (49), 120 (61), 119 (100), 107 (87), 106 (68), 94 (40), 93 (67), 92 (40), 82 (31), 81 (32), 69 (41), 67 (34), 59 (37), 55 (32), 41 (69), 39 (65). HR-MS: 208.1453 (C₁3H₁₆²H₂O₂, M^{+} , calc. 208.1432).

 $(8R,6Z,9Z)-[7,8-^2H_2]$ Trideca-6,9-dien-3-ynoic Acid ((8R)-10). Prepared from (8R)-9 (0.48 g, 2.5 mmol) as described for (8S)-10. Yield: 0.32 g (64%). Spectroscopic data: identical with those of (8S)-10.

 $(8S, 3Z, 6Z, 9Z)-[7, 8-^2H_2]$ Trideca-3,6,9-trienoic Acid ((8S)-11). A soln. of (8S)-10 (0.32 g, 1.5 mmol) in dry THF (10 ml) is hydrogenated over Lindlar's catalyst (100 mg, Fluka, Buchs, Switzerland) in the usual manner. When the uptake of H₂ ceases, the catalyst is filtered off, the solvent evaporated, and the crude acid purified by HPLC (*RP* 18, MeOH/H₂O 4:1): 0.29 g (90%). IR (film): 3600–3200s (br.), 3014m, 2964s, 2932m, 2875m, 2346w, 1711s, 1414w, 1263m, 1218w, 1162m, 1115m, 933w, 864w, 796w, 692w, 662w. ¹H-NMR (CDCl₃/²H₂O, 400 MHz): 5.60–5.34 (*m*, 5 olef. H); 3.18 (*d*, 2 H–C(2)); 283 (*t*, 2 H–C(5)); 2.8 (br. *d*, 1 H–C(8)); 2.02 (*q*, 2 H–C(11)); 1.37 (*sext.*, 2 H–C(12)); 0.9 (*t*, 3 H–C(13)). MS (methyl ester; 70 eV): 193 (11, M^{++} – OMe), 177 (5), 175 (6), 165 (7), 163 (5), 151 (33), 150 (31), 137 (12), 135 (21), 123 (20), 121 (37), 111 (35), 107 (62), 97 (40), 95 (57), 82 (66), 81 (92).

761

80 (64), 69 (80), 68 (63), 59 (35), 55 (39), 41 (97), 39 (100). HR-MS (free acid): $210.1601 (C_{13}H_{18}^{-2}H_2O_2, M^{++}, calc. 210.1588).$

 $(8 \text{ R}_{3} \text{ Z}_{6} \text{ Z}_{9} \text{ Z}_{-} \text{ / } 7^{8-2} \text{ H}_{2}]$ Trideca-3,6,9-trienoic Acid ((8*R*)-11). Prepared from (8*R*)-10 (0.30 g, 1.44 mmol) as described for (8*S*)-11. Yield: 0.27 g (89%). Spectroscopic data: identical with those of (8*S*)-11. HR-MS (methyl ester): 224.1733 (C₁₄H₂₀²H₂O₂, *M*⁺, calc. 224.1745).

 $(5 \text{ S}, 3 \text{ Z}, 6 \text{ Z}) - [4, 5^{-2}H_2]$ Deca-3,6-dien-1-ol ((5S)-12). At -78° , (2S)-8 (0.56 g, 4.42 mmol) is added to a stirred soln. of $\{3\text{-}[(\text{tetrahydro-}2H\text{-pyran-}2\text{-y}])$ oxy] propylidene $\}$ triphenyl phosphorane (prepared at 0° from the corresponding phosphonium bromide (3.4 g, 6.9 mmol) by addition of 1 equiv. of BuLi) in dry THF (100 ml). The soln. is allowed to come to r.t., and the product is isolated, deblocked, and purified as described for (8S)-9. Chromatography on silica gel furnished 0.45 g (61%) of (5S)-12. Colorless oil. IR (film): 3345m (br.), 3013m, 2962s, 2933s, 2874s, 2233w, 1648w, 1461m, 1377w, 1233w, 1111w, 1049s, 902w. ¹H-NMR (CDCl₃, 250 MHz): 5.44–5.27 (m, 3 olef. H); 3.63 (t, 2 H–C(1)); 2.77 (br. s, 1 H–C(5)); 2.35 (q, 2 H–C(2)); 2.03 (q, 2 H–C(8)); 1.51 (br. s, OH); 1.39 (sext., 2 H–C(9)); 0.89 (t, 3 H–C(10)). MS (70 eV): 156 (0.5, M⁺⁺), 138 (4), 123 (9), 109 (13), 95 (38), 81 (92), 80 (77), 69 (100), 68 (67), 56 (43), 55 (38), 41 (92), 39 (80).

 $(5 \text{ R}_3 \text{ }_2 \text{ }_5 \text{ }_2 \text{ }_1 \text{ }_4 \text{ }_5 \text{ }_2 \text{ }_1 \text{ }_2 \text{$

 $(5S, 3Z, 6Z) - [4, 5 - ^{2}H_{2}]$ Deca-3,6-dienoic Acid ((5S)-13). Alcohol (5S)-12 (0.30 g, 1.92 mmol) is oxidized with Cr(VI) and purified as described for (8S)-10. Yield: 0.196 g (60%). IR (film): 3600-3200m (br.), 3015m, 2964s, 2936s, 2874m, 2236w, 1711s, 1457w, 1414m, 1377w, 1294m, 1217m, 1171m, 1111w, 932w. ¹H-NMR (CDCl₃, 250 MHz): 5.56 (t, J = 7.2, 1 H-C(3)); 5.46-5.28 (m, 2 olef. H); 3.16 (d, 2 H-C(2)); 2.77 (br. s, 1 H-C(5)); 2.05 (q, 2 H-C(8)); 1.40 (sext., 2 H-C(9)); 0.90 (t, 3 H-C(10)). MS (70 eV): 170 (4, M^{++}), 152 (8), 137 (5), 133 (5), 123 (8), 122 (8) 110 (19), 109 (21), 95 (28), 81 (96), 80 (73), 69 (81), 68 (62), 55 (41), 45 (79), 41 (100), 39 (89). HR-MS: 170.1263 (C₁₀H₁₄²H₂O₂, M^{++} , calc. 170.1275).

 $(5R,3Z,6Z)-[4,5-^2H_2]$ Deca-3,6-dienoic Acid ((5R)-13). Prepared from (5R)-12 (0.30 g, 1.92 mmol) as described for (5S)-13. Yield: 0.18 g (55%). Spectroscopic data: identical with those of (5S)-13.

REFERENCES

- [1] W. Boland, K. Mertes, Eur. J. Biochem. 1985, 147, 83.
- [2] F. Bohlmann, C. Zdero, D. Berger, A. Suwita, P. Mahanta, C. Jeffrey, Phytochemistry 1979, 18, 79.
- [3] R. L. Maas, C. D. Ingram, A. T. Porter, J. A. Oates, D. F. Taber, A. R. Brash, J. Biol. Chem. 1985, 260, 4217.
- [4] E.J. Corey, P.T. Lansbury, J. Am. Chem. Soc. 1983, 105, 4093.
- [5] L. Brandsma, in 'Preparative Acetylenic Chemistry', Elsevier, Amsterdam, 1971, p. 169.
- [6] R. Julina, T. Herzig, B. Bernet, A. Vasella, Helv. Chim. Acta 1986, 368.
- [7] J.A. Dale, D.L. Dull, H.S. Mosher, J. Org. Chem. 1969, 34, 2543.
- [8] Y. Gao, R. M. Hanson, J. M. Klunder, S. Y. Ko, H. Masamune, K. B. Sharpless, J. Am. Chem. Soc. 1987, 5765.
- [9] W. A. König, S. Lutz, G. Wenz, G. Görgen, C. Neumann, A. Gäbler, W. Boland, Angew. Chem. 1989, 101, 180.
- [10] L. X. Dai, B. L. Lou, Y. Z. Zhang, G. Z. Guo, Tetrahedron Lett. 1986, 27, 4343.
- [11] C. A. Brown, V. K. Ahuja, J. Org. Chem. 1973, 38, 2226.
- [12] C. Neumann, W. Boland, Eur. J. Biochem., in press.